

Hypolipidemic Effect of Apigenin Extracted From Parsley (*Petroselinum sativum* L.) Leaves In Cadmium Chloride Treated Rats (Part II)

*Khalisa Khadhim Khudiar

**Aous Khahtan Ahmed Al-Mezain

***Khatan Ahmed Al-Mezain

*Department of Physiology and Pharmacology / College of Veterinary Medicine /Baghdad University-Iraq.

** Iraqi Center For Cancer and Medical Genetics Researches, Al-Mustansiriya University-Iraq.

*** Department of pathological analysis techniques/ Dijlah University College / Baghdad -Iraq.

Corresponding author: *E.mail:* Khalisakhadim0@gmail.com

Abstract

Cadmium has been reported to have cumulative effects on mortality ,cardiovascular , renal and neurologic .flavonoids (apigenin) are naturally occurring phytochemicals possessed divers pharmacological effects hypolipidemic and antioxidant activity .this study was carried out to investigate the protective effect of the flavonoids extracted from (*Petroselinum sativum*) parsley leaves on some parameters related to cardiac risk in adult male rats exposed to 50 ppb cadmium chloride in drinking water . Crude flavonoids were extracted from parsley leaves . Further purification of flavonoids was performed by gel permeation column chromatography (TLC) . Thirty adult Albino male rats randomly and equally divided into three group (10 rats for each) and were treated daily for 60 days as follows : The rats were kept on ordinary tap water as control (group c) , received drinking water containing 50 ppb of cadmium chloride (group T1) and simultaneously given orally 150mg/kg B.W. of flavonoids (apigenin) extracted from parsley in addition to 50ppb of cadmium chloride in drinking water (group T2). Fasting blood sample were collected by cardiac puncture technique at 0,30 and 60 days of experiment for measuring the flowing parameters : serum lipid profile including: a. triacylglycerol (TAG) .b. serum total cholesterol (TC) .c. serum high density lipoprotein (HDL-C). d. serum low density lipoprotein (LDL-C).e. serum very low density lipoprotein (VLDL-C) concentration .Besides section from heart, aorta were taken at the end of experiment for histopathological study .The results showed that the yield of crude flavonoids from parsley leaves were approximately 2.68% of dry leaves .Purification of crude flavonoids on Sephadex LH-20 clarified three peaks activity and the proportions of the purified fraction P1, P2 and P3 were 14.06, 82.43 and 3.5% respectively. Thin layer chromatography confirmed that p2 was pure apigenin . the result revealed that animal exposed to 50 ppb of cdcl2 water for 60 days caused cardiac damage manifested by a significant elevation in serum TC,TAG,HDL-C,VLDL-C and LDL-C concentrations, with a significant depression in serum HDL-C concentration . Gavage of flavonoids and Cd concurrently caused a significant correction of the previous studies parameters . Histological section of the heart and aorta of cd treated (T1) group revealed an occurrence of early signs of atherosclerotic lesions . Such histological change could not be observed in heart and aorta of animal in group

T2 after oral intubation of flavonoids (apigenin). It could be concluded that apigenin is effective in prevention the deleterious effect of cadmium chloride on major risk factors of cardiovascular system.

Key Words: Cadmium chloride, apigenin, lipid profile, heart, parsley.

التاثير الخافض للدهون للابجينين المستخلص من اوراق نبات المعدنوس في الجرذان المعاملة بكلوريد الكاديوم

*خالصه كاظم خضير ، **اوس قحطان احمد المزين *** قحطان احمد المزين

*فرع الفسلج والأدوية /كلية اطب البيطري -جامعة بغداد-العراق

**المركز العراقي لبحوث السرطان والوراثة الطبيه - الجامعة المستنصرية-العراق

***قحطان احمد المزين / قسم تقنيات التحليلات المرضيه - كلية دجلة الجامعة- العراق

المستخلص

اجريت هذه الدراسة لمعرفة الدور الوقائي القلبي للفلافونيدات المستخلصه من اوراق البقدونس في ذكور الجرذان البالغة المعرضه لكلوريد الكاديوم 50 جزء بالبلليون في ماء الشرب . تم استخلاص الفلافونيدات الخام من اوراق البقدونس ، ثم تنقيه (الابجينين) على هلام عمود التحلل الوني على السيفادكس LH-20 ومن ثم تشخيصه باستخدام كروموتورفيا الطبقة الرقيقه . وقد بلغت كميته المستخلص الخام من الفلافونيدات 2.68 غم لكل 100 غم من الاوراق الجافه ، ووضحت نتائج التنقيه على عمود التحلل الوني وجود ثلاث قيم بتركيز 14.06,82.43,3,51% وقد اكدت نتائج الكوموتوكرافيا الطبقة الرقيقه ان P2 هو الابجينين الذي تمت تنقيته وقد بلغت تركيز الابجينين النقي 47% من الاوراق الجافه للبقدونس . تم تقسيم 30 من ذكور الجرذان البالغة عشوائيا الى ثلاثة مجاميع (عشر حيوانات/مجموعه) وعوملت كلتالي لمدته 60 يوما : اعطيت المجموعه الاولى الناء العادي واعدت مجموعته سيطره (Group C)، في حين اعطيت حيوانات المجموعه الثانيه ماء شرب محتوي على 50 جزء بالبلليون من كلوريد الكاديوم (Group T1) ، اما حيوانات المجموعه الثالثه فقد اعطيت ماء الشرب المحتوي على 50 جزء بالبلليون من كلوريد الكاديوم بالاضافه الى تجريع الفموي للفلافونيدات المستخلصه من اوراق البقدونس بتركيز 150 ملغم/كغم من وزن الجسم (Group T2) ، تم جمع عينات الدم الوخز القلبي في الفترات 0 و 30 و 60 يوما من التجربه لغرض قياس تركيز الكولسترول الكلي (TC) والكليسيريدات الثلاثيه (TAG) والكولسترول في البروتينات الشحميه ذات الكثافه العاليه (HDL-C) والواطئه (LDL-C) والواطئه جدا (VLDL-C) بالاضافه الى اخذ مقاطع نسيجييه للابهر والعضله القلبيه . اظهرت نتائج هذه الدراسة ان تعرض الحيوانات الى كلوريد الكاديوم بتركيز 50 جزء بالبلليون في ماء الشرب لمدته 60 يوما قد تسبب حدوث تلف في القلب تمثل بزياده معنويه في (TC,LDL-C,VLDL-C) . بينما ساهم تجريع الفلافونيدات مع الكاديوم (T2) في حصول تغيرات ايجابيه معنويه في المعايير التي اشير اليها سابقا والتي تمثلت بحصول زياده معنويه في تركيز الHDL-C،بالاضافه الى حصول انخفاض معنوي في تراكيز كل من VLDL-C ,LDL-C,TC,TAG

في مصل الدم . اظهرت نتائج المقاطع النسيجية للقلب والابهر في المجموعه المعرضه للكاديوم T1 في وجود مؤشرات اوليه لافه التصلب العصيدي تجلت بصوره رئيسيه بارتشاح الخلايا الالتهابيه (الخلايا العدله والالتهابيه) على جدار الابهر وارتشاح الدهون في الطبقة المصليه بالاضافه الى ارتشاح خلايا وحيدة النواه في منطقه ال intima layer بينما ادت المعامله بالفلافونيدات المستخلصه من اوراق البقدونس (T2) الى قله ظهور التلف في القلب . لقد اكدت نتائج هذه الدراسه التاثير الوقائي والدور المانع للاكسده للفلافونيدات المستخلصه من اوراق البقدونس ضد التلف الحاصل في القلب تحت تاثير الكاديوم .

الكلمات المفتاحيه : كلوريد الكاديوم ، اجنين ، صورة الدهون ، القلب ، المعدنوس .

Introduction

Cadmium (Cd) is an environmental pollutant that is released naturally from minerals ,forest fires and volcanic emission (12) cigarette smoke ,tap and well water ,food ,fungicides and seafood are regarded as important sources of pollution with Cd (4,15) .Besides Cd is a product of zinc and lead mining and smelting ,which are important sources of environmental pollution (11). On the other head , a major role of cd intake (for nonsmoker) is ingested ,this is largely attributed to the presence of trace level of cd in food stuff of natural origin e.g. cereals ,beans ,carrots, tomatoes , beverage coffee and tea (34,50). This heavy metals is now have numerous undesirable effects on health in experimental animals and humans (7) , including kidney (51), bone (24), liver(54) and nervous system (27) . It has been suggested that the exposure of cd has been associated with wide variety of cardiovascular disease such as atherosclerosis (36),heart failure (46) and cardiomyopathy (3,41). Flavonoids are a diverse group of low molecular mass polyphenolic compounds widely distributed in plants ,they occur naturally in broad range of fruits ,vegetable ,nuts ,seeds ,herbs, spices, stems ,flowers and beverages such as green tea and red wine (25,32,52,53) ,there are more than 8000 different compounds (58). Addition to its effects on reproductive system (18), flavonoids have been reported to exert multiple biological effects , including antioxidant, free radicals scavenging abilities (19) , diuretics (8), anti-inflammatory and anti-carcinogenic activity (9).Apigenin is natural flavonoids present abundantly in common fruits and vegetable such as parsley (30,33,42) ,onion , orange ,tea, chamomile wheat, sorouts, apples broccoli(16,20). This study was undertaken to assess the relative efficacy of apigenin extracted from parsley on some biomarkers related to cardiac functions of cadmium treated rats.

Materials and methods

Extraction of flavonoids from parsley (*Petroselinum sativum*) leaves

Crude flavonoids were extracted from parsley leaves according to the method of harbore (17) modified by AL-Kawary (2). Then extracted flavonoids were subjected to further purification through solubilization in 85% ethanol and gel filtration on Sephadex LH-20 . Dried flavonoids fraction then was subjected to further analysis using thin layer chromatography technique.

Experimental Design:

Thirty adult (between 2.5-3 months) Albino male rats (175-225gm) were randomly and equally divided into three groups (10 rat/ group) and were treated daily for 60 days as follows: 1. Group C (control). 2. Group T1: Rats of these group were allowed to ad libitum supply of drinking water containing 50 ppb of CDCL2 .3. Group T2: Rats of these group were received 150mg/kg B.W. of flavonoids (apigenin) extracted from parsley orally, in addition to 50ppb of cdcl2 in drinking water . fasting blood samples were collected by cardiac puncture technique at 0,30,60 days of experiment for measuring the following parameters .Serum (TAG) ,(TC) and (HDL-C) using enzymatic kit (linear chemicals Barcelona, spian) , (LDL-C) and (VLDL-C) concentration (13). Besides, sections from heart and aorta were taken at the end of experiment for histological study (26). Statistical analysis of data was performed on the basis of Two –Way Analysis of Variance (ANOVA) using SAS® software (SAS, 13) at a significant level of (P< 0.05). Differences were determined using least significant differences (LSD) (55).

Results

The yield of crude flavonoids extracted from samples revealed that out of each 100 gm. of dry ground leaves parsley was approximately 2.68 g. of crude flavonoids was obtained .Three peaks were found in the supernatant of the extract (figure-1), (p1,p2 and p3) on UV 330 nm and the proportion of these peaks were 14.06, 83.43 and 3.5% respectively .The major peak (p2) was collected and dried under vacuum then subjected to thin layer chromatography analysis on silica gel (figure-2) ,and the RF of purified fraction was 0,81 . Such value was approximately the same of apigenin as recorded by Harborne (17), under similar diagnostic conditions.

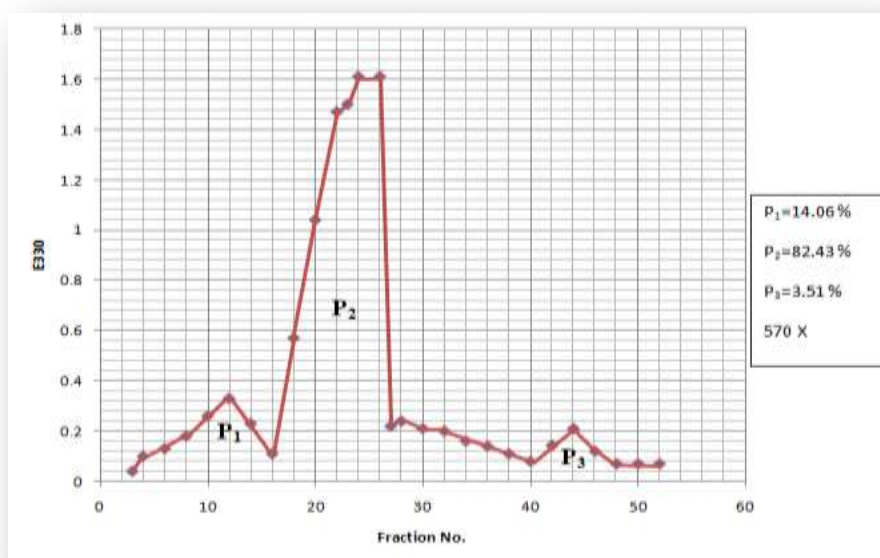


Figure -1 : Purification of crude flavonoids from parsley on Sephadex LH-20 column (38 × 1.6) ID. Ethanol (85 %) was used as an eluent at flow rate of 30 ml/hr. , fraction size: 4 ml.



**Figure -2: TLC analysis of flavonoids extracted by Harborne (1973).
1. Purified apigenin on Sephadex LH-20. 2. Crude apigenin .**

Lipid profile testes:

Serum TC, TAG, LDL-C and VLDL-C concentration showed a significant increase ($p < 0.05$) at the day 60 of treatment (tables 1-4) in rats received 50ppb of cdcl4 compared with control and T2 groups where oral intubation of extracted flavonoids (T2group) significant suppressed ($p < 0.05$) the elevated previously mentioned parameters and the values tend to normalize that of the control. On the other hand a significant increase ($p < 0.05$) in serum HDL-C concentration (table 5) was recorded in T2 group as compared To the T1 treated group at the san=me treated period (after 60 days) .

Table -1: Effect of flavonoids extracted from (*Petroselinum sativum*) leaves on serum total cholesterol (mg/dl) concentration in control and cadmium treated rats.

Groups Days	C	T1	T2
Zero	93.8 ± 3.10 A a	94.5 ± 2.41 A a	96.5 ± 2.41 A a
30	93 ± 3.12 A a	95.5 ± 3.38 A a	91.6 ± 2.05 A a
60	95.6 ± 3.04 A a	110.3 ± 3.56 B b	94.1 ± 3.18 A a

Values are expressed as mean ± SE, n = 6 each group

C: control group. T1: Animals received 50 ppb of CdCl₂ in drinking water.

T2: Animals received 50ppb of CdCl₂ and 150mg/kg B.W. of flavonoids extracted from parsley.

Capital letters denote differences between groups, P<0.05 vs. control.

Small letters denote differences within group, P<0.05 vs. control.

Table-2: Effect of flavonoids extracted from (*Petroselinum sativum*) leaves on serum triacylglycerol (mg/dl) concentration in control and cadmium treated rats.

Groups Days	C	T1	T2
Zero	87.3 ± 3.54 A a	88.3 ± 4.01 A a	89.6 ± 3.14 A a
30	88.5 ± 4.34 A a	89.8 ± 3.59 A a	84.8 ± 2.42 A a
60	88.8 ± 3.37 A a	105.3 ± 3.06 B b	86.6 ± 3.15 A a

Values are expressed as mean ± SE, n = 6 each group

C: control group. T1: Animals received 50 ppb of CdCl₂ in drinking water.

T2: Animals received 50ppb of CdCl₂ and 150mg/kg B.W. of flavonoids extracted from parsley.

Capital letters denote differences between groups, P<0.05 vs. control.

Small letters denote differences within group, P<0.05 vs. control.

Table-3: Effect of flavonoids extracted from (*Petroselinum sativum*) leaves on serum low density lipoprotein-cholesterol (LDL-C) (mg/dl) concentration in control and cadmium treated rats.

Groups Days	C	T1	T2
Zero	42.4 ± 3.17 A a	41.3 ± 5.22 A a	44.4 ± 1.48 A a
30	40.9 ± 3.55 A a	45.7 ± 5.22 A a	41.6 ± 1.94 A a
60	42.4 ± 2.31 A a	64.4 ± 5.11 B b	45.6 ± 3.93 A a

Values are expressed as mean ± SE, n = 6 each group

C: control group. T1: Animals received 50 ppb of CdCl₂ in drinking water.

T2: Animals received 50ppb of CdCl₂ and 150mg/kg B.W. of flavonoids extracted from parsley.

Capital letters denote differences between groups, P<0.05 vs. control.

Small letters denote differences within group, P<0.05 vs. control.

Table -4: Effect of flavonoids extracted from (*Petroselinum sativum*) leaves on serum very low density lipoprotein-cholesterol (VLDL-C) (mg/dl) concentration in control and cadmium treated rats.

Groups Days	C	T1	T2
Zero	17.4 ± 0.71 A a	17.7 ± 0.79 A a	17.9 ± 0.63 A a
30	17.7 ± 0.86 A a	17.9 ± 0.77 A a	16.9 ± 0.44 A a
60	17.7 ± 0.67 A a	21.06 ± 0.61 B b	17.3 ± 0.63 A a

Values are expressed as mean ± SE, n = 6 each group

C: control group. T1: Animals received 50 ppb of CdCl₂ in drinking water.

T2: Animals received 50ppb of CdCl₂ and 150mg/kg B.W. of flavonoids extracted from parsley.

Capital letters denote differences between groups, P<0.05 vs. control.

Small letters denote differences within group, P<0.05 vs. control.

Table-5: Effect of flavonoids extracted from (*Petroselinum sativum*) leaves on serum high density lipoprotein-cholesterol (HDL-C) (mg/dl) concentration in control and cadmium treated rats.

Groups Days	C	T1	T2
Zero	34 ± 1.48 A a	35.5 ± 1.73 A a	34.16 ± 0.87 A a
30	32.6 ± 1.33 A a	31.8 ± 1.58 A a	33.2 ± 2.33 A a
60	35.5 ± 1.84 A a	24.8 ± 1.74 B b	30.6 ± 2.04 C a

Values are expressed as mean ± SE, n = 6 each group

C: control group. T1: Animals received 50 ppb of CdCl₂ in drinking water.

T2: Animals received 50ppb of CdCl₂ and 150mg/kg B.W. of flavonoids extracted from parsley.

Capital letters denote differences between groups, P<0.05 vs. control.

Small letters denote differences within group, P<0.05 vs. control.

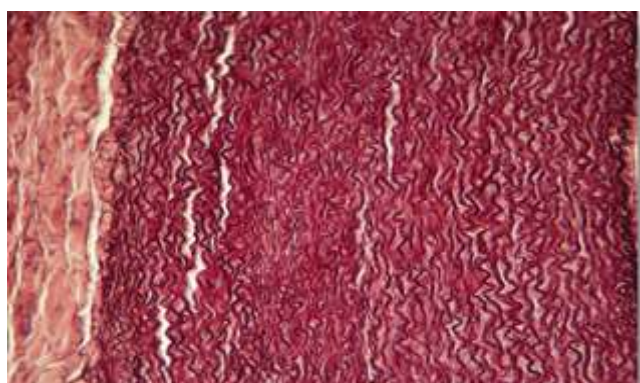


Figure -3: Histological section of normal aorta of rat (H and E, X40).

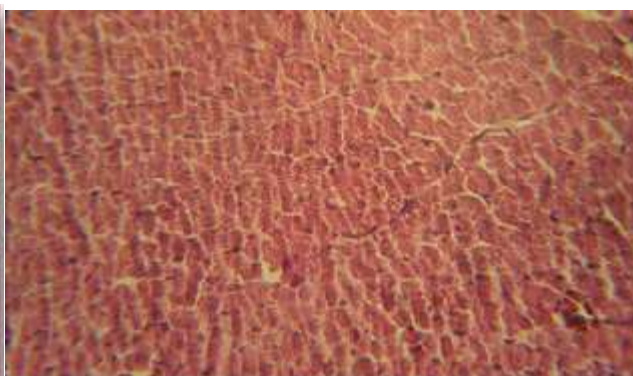


Figure -3: Histological section of normal aorta of rat (H and E, X40).

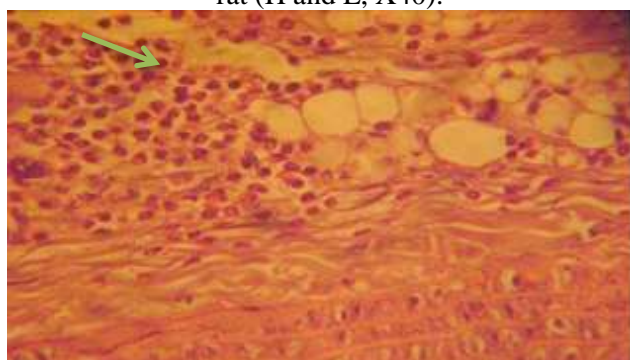


Figure-5: Histological section of aorta of rat treated with CdCl₂ (50 ppb) in drinking water (T1). Note: severe inflammatory cell infiltration mainly, neutrophil and macrophage around the aorta (arrow). (H and E, X40)

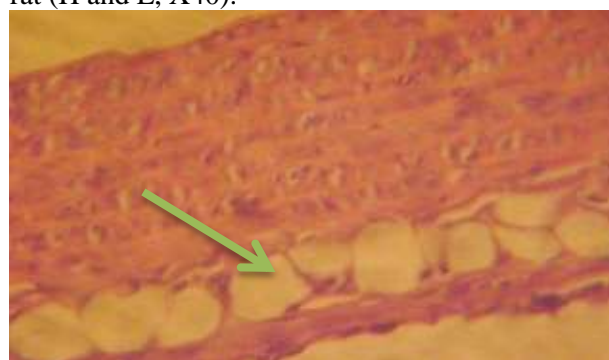


Figure-6: Histological section of aorta of rat treated with CdCl₂ (50ppb) in drinking water (T1). Note: fatty infiltration in serosal layer (arrow). (H and E, X40).

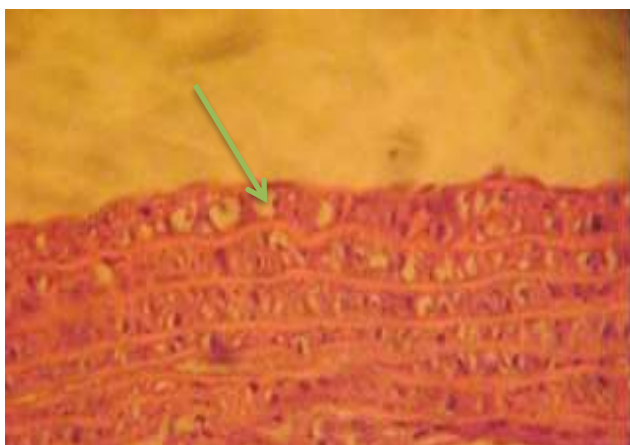


Figure -7: Histological section of aorta of rat treated with CdCl₂ (50ppb) in drinking water (T1). Note: vacuolation on mononuclear cell infiltration in the intima of the aorta (arrow). (H and E, X40).

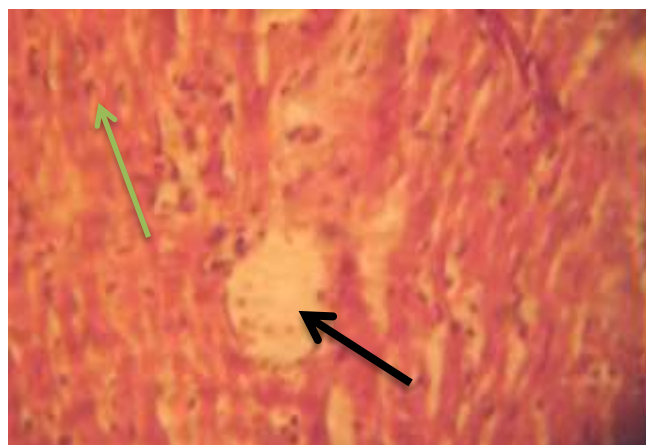


Figure-8: Histological section of heart of rat treated with CdCl₂ (50ppb) in drinking water (T1). Note: inflammatory cell infiltration between the muscle fiber (→) and congestion of blood vessels (→) (H and E, X40).

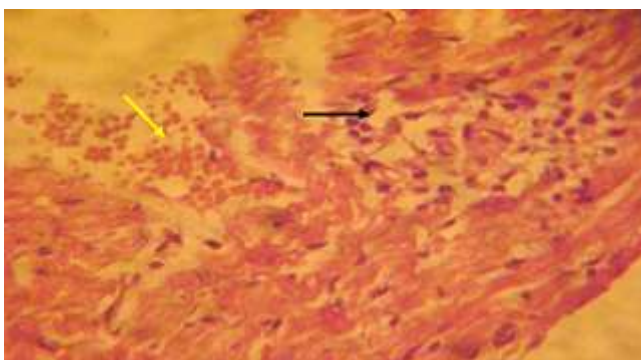


Figure-9: Histological section of heart of rat treated with CdCl₂ (50ppb) in drinking water (T1). Note: Inflammatory cell infiltration mainly neutrophil and macrophage in the atrium (→), and congestion of blood vessels (→). (H and E, X40).

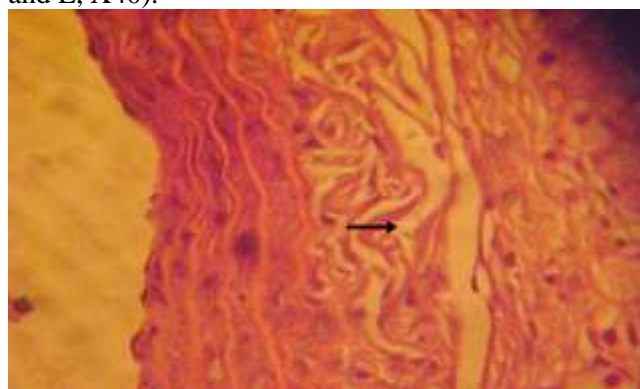


Figure-10: Histological section of aorta of rat treated with CdCl₂ (50ppb) in drinking water plus flavonoids 150 mg/kg B.W. (T₂). Note: partial regression of the lesion with few inflammatory cell infiltration between the muscle fibers (arrow) (H and E, X40).

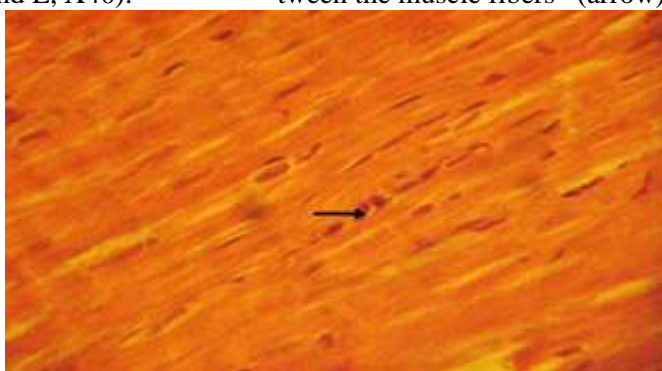


Figure -11: Histological section of heart of rat treated with CdCl₂ (50ppb) in drinking water plus flavonoids 150 mg/kg B.W. (T₂) Note: there is few inflammatory cells infiltration between the muscle fiber (arrow) (H and E, X40).

Discussion:

Hyperlipidemic effect of cadmium is agreement with (14,39,44,47) . The observed dyslipidemia after Cd exposure may reflect the suppression of lipid metabolism and reduction in antioxidant enzymes duo to Cd induced oxidative stress Disturbing the antioxidant defense system after Cd exposer as result of increase quenching enormous free radicals (FRs) produced under such condition (22,40,49) may be responsible for such depression in HDLC concentration and elevation in LDL-C (10). This suggestion is documented in our previous study (23), where significant decrease in GSH (non-enzymatic antioxidant) and elevation in MDA concentration were observed after Cd exposure. Olisekodiaka and his colleagues,(38) confirmed that Cd can inversely affect lipoprotein and lipid profile via lipid peroxidation.

The results also explained the lowering effect of parsley extract on TC, TAG, LDL-C concentration , which was documented in vivo (21,56,57). Parsley hypolipidemic effects may be attributed to its flavonoids content , mainly apigenin (42), that play a role in lowering activity of both HMG-CoA reductase and acetylCoA cholesterol-o-acetyl transferase (ACAT) (31), Flavonoids decrease activity of rate limiting enzyme of cholesterolgenesis by phosphorylation of enzyme directly (60), leading to hypercholesterolemia . Besides, flavonoids possess beneficial effect on cardiovascular risk factor such as lipoprotein oxidation , dyslipidemia, endothelial dysfunction and regulation of serum lipid profile (28). As well as , the free radicals scavenging activities of parsley (29) and apigenin (43) may reduce LDL-C oxidation and depressed its transportation to different tissues.

Elevation in HDL-C concentration after parsley extract , could be due to stabilizing effect of its polyphenols on plasma lipoprotein or due to systemic effect of flavonoids to modulate various enzymatic activity that can affect lipoprotein leading to augmentation of HDL-C. Such elevation in HDL-C is in agreement with (1,5) , where increase HDL-C facilitate transport of cholesterol from serum to liver, where its catabolized and excreted from bodies (59). Besides high HDL-C concentration is correlated with decrease risk of cardiovascular disease (48).

Histopathological observation of aorta and heart tissue of animal in group T1 (figures-3,4,5) also confirmed the cardio toxic effect of CdCl₂ (35,45) comparing to control (figure-2). Sections of muscle (figures-6,7) manifested by occurrence of atheromatous lesion indicating injury of vascular endothelium after cadmium exposure (46). Which may be due to the function role of hydrogen radical generation in the endothelium (37) .These radical have been documented to be responsible for endothelial dysfunction and atheroma (6). The present study also confirm the curative effect of apigenin which causes regeneration of cardiac cell(figure-8,9). This may be due to its antioxidant properties .

References

- 1- Abd El-Baky, A.E. (2011). Quercetin Protective Action On Oxidative Stress, Sorbitol, Insulin Resistance And β -Cells Function In Experimental Diabetic Rats. International Journal Of Pharmaceutical Studies And Research., 2(2):11-18.
- 2- Alkawary, T.A.A. (2000). Extraction Of Some Flavonoids From *Sisyrinchium spian-chisti* And Its Antioxidant Effects In Sunflower Oil. Ph.D. Thesis. College Of Agriculture – University Of Baghdad- Iraq.
- 3- Al-Okaily, B .N.; Salman, A.D. And Khudiar, K.K .(2013) . The Ameliorative Effect Of Vitamin E On Electrocardiogram Of Rabbits Exposed To Cadmium Chloride . MRVSA 2(1), 24-35. [Http://Mirror Of Research in veterinary sciences and animals.Com/](http://MirrorOfResearchinveterinarysciencesandanimals.Com/) ISSN 2307-8073.
- 4- Asagba S.A; Eriyamremu, G.E. And Igberaese, M.E.(2008). Bioaccumulation Of Cadmium And Its Biochemical Effect On Selected Tissue Of The Catfish (*Clarias Gariepinus*) .Fish Physiol.Biochem.,34:61-60.
- 5- Baghdadi , H.H.(2014). Antioxidant Potential Of Quercetin: Remarkable Protection Against Hypercholesterolemia In Rats. British Journal Of Medicine And Medical Research ., 4(26):4382-4391.
- 6- Berndt, C; Lillig, C.H. And Holmgren, A. (2007). Thiol-Based Mechanism Of The Thioredoxin And Glutaredoxin System : Implications For Disease Of The Cardiovascular System. Am J. Physiol. Heart Circ. Phy., 292, P: H1227, H1236.
- 7- Cannon ,G; Ferruggia, E; Luparello, C. And Rinaldi A.(2009) Cadmium And Mitochondrial. Mitochondrion,9:377-384.
- 8- Decampos, K.E.D; Balbi, A.P.C. And Alves, M.J.Q.D.F.(2009)_ . Diuretic And Hypotensive Activity Of Aqueous Extract Of Parsley Seeds (*Petroselinum Sativum Hoffm*) . In Rats .Brazilan. J Pharmacol., 19(A1):41-45.
- 9- Deendayal,P; Sanjeev, S. And Sanjay , G. (2007). Apigenin And Cancer Chemoprevention : Progress, Potential And Promise (Review) Inter. J. Of Ocol., 30:233-245.
- 10- Djukić-Ćosića, D.; Jovanovića, M.C.; Bulata,Z. P.; Ninkovićb,M.; Maličevićb,Z. And Matovića,V. (2008). Relation Between Lipid Peroxidation And Iron Concentration In Mouse Liver After Acute And Subacute Cadmium Intoxication. J Trace Elements Med Biol, 22, (1) 19 :66–72.
- 11- Duruibe, J.O.;Ogwuegbu, M.O.C. And Egwurugwu , J.N.(2007). Heavy Metal Pollution And Human Biotoxic Effects, Int. J Phys. Sci.2(5):112-118.
- 12- Egan K, Hamridge T, Kayama F. Cadmium: WHO Food Additive Series. Geneva: 2005. Cadmium Impact Assessment Of Different Maximum Limits; Pp. 35–46.
- 13- Friedewald, W; Levy, Y. And Fredrickson (1972). Estimation Of The Concentration Of Low- Density Lipoprotein Cholesterol In Plasma Without Use Of Preparative Ultracentrifuge .Clin .Chem, 18:499-502.

- 14- Ghosh K. And Indra N (2015). Research Article Hypoglycemic And Hypolipidemic Potential Of *Centella Asiatica* Ethanolic Extract On Cadmium Intoxicated Albino Rats. *Inter Jrecent Scient Res.*, 6 (7):5327-5332.
- 15- Germolec, D.R; Yang, R.S.; Acermann, M.F. Abd Luster, M.I.(2002). Toxicological Studies Of Chemical Mixture Of 25 Ground Water Contaminants .II Immunosuppression In B6C3FI Mice. *Fundan. Appl. Toxicol.*,13:377-387.
- 16- Halvorsen, B; Hotel ,K And Myhestad, M.(2002).A Systemic Screening Of Total Antioxidant In Dairy Plants . *J. Nutr.*, 132:461-471.
- 17- Harborne ,J.B. (1984). *Phytochemical Methods .Aguide To Modern Techniques Of Plant Analysis.*London Chapman And Hall Ltd., Pp 49-188.
- 18- Hozayen, W,G.; Hegab, M.Y. And Soliman, H.A..(2015). Effects Of Parsley And Pumpkin On Alcohol Induced Testicular Damage In Rat Model . *J. Int. Academic Res. For Multidisciplinary.*, Volume 2, Issue 12, Www.Jiarm.Com .
- 19- Ivanova, J.; Gluhcheva, Y.; Kamenova, K.; Arpadian, S. And Mitewa, M. (2014). Monensin Ameliorates Cadmium-Induced Hepatic Injury In Mice, Subjected To Subacute Cadmium Intoxication. *J Biotechnol Equip*, 28(1): 147-152.
- 20- Janssen,K; Mensink, R.P; Cow. F.J. (1998). Effects Of Flavonoids Quercetin And Epignin On Hemostasis In Healthy Volunteers : Results From An In Vitro And Dietary Supplement Study .*Am J Clin Nutr.*, 67:255-262.
- 21- Khalil ,A.F.; Elkatry, H.O. And Mehairy, H.F. (2015). Protective Effect Of Peppermint And Parsley Leaves Oils Against Hepatotoxicity On Experimental Rats. *Annals Agri Sci.*, 60(2) :353–359.
- 22- Khudiar , K.K; Abudulla, B.N. And Al-Mzaien, K.A.(2001). The Protective Effect Of Aqueous Extract Of Parsley *Petroselinum Sativum* Seeds On Experimentally Induced Oxidative Stress In Rats. *IJVM.* 25(1): 153-171.
- 23- Khudair, K.K. And Ahmed, A.K. (2012).Protective Effects Of Flavoroids Extracted From Parsley (*Petroselinum Sativum* L-) Leaves On Liver Function In Male Rats Exposed To Cadmium Chloride. *Iraqi J Biotechnol*,11(1) : 90-108.
- 24- Lavery ,T.J; Kemper, C.M; Sanderson, K;Schultz, C.G; Coyle, P; Mitchell, J.C. And Seuront, L.(2009). Heavy Metal Toxicity Of Kidney And Bone Tissue In South Australian Adultbottlenose Dolphins (*Tursiops Aduncus*). *Mar Environ Res.*, 67(1): 1-7.
- 25- Lee, K.W; Kim, Y.J; Kim, D.O; Lee, H.J. And Lee, C.Y.(2003). Major Phenolics In Apple And Their Contribution To The Total Antioxidant Capacity. *J Agri Food Chem.*, 51(22): 6516-6520.
- 26- Luna, L.G. And Lee,G. (1968). *Manual Of Histological Staining Methods Of Armed Forces Institutes Of Pathology.* 3rded .Mc Grow-Hill Book Company .New York. Pp:12-31.

- 27- Maodaa,S.N.; Allam,A.A.; Ajarem,J.; Abdel-Maksoud,M.A.; Al-Basher, G.I. And Zun Yao Wang, Z. Y.(2016). Effect Of Parsley (*Petroselinum Crispum*, Apiaceae) Juice Against Cadmium Neurotoxicity In Albino Mice (*Mus Musculus*). Behavioral And Brain Functions., 12:6 . DOI: 10.1186/S12993-016-0090-3.
- 28- Mahmoud, K.A.(2011). Antidiabetic And Antioxidant Effects Of Parsley Extract (*Petroselinum Crispum*) On Diabetic Rats. Isotope Rad Res, 43:341–57.
- 29- Mahmood S, Hussain S, Malik F.(2014). Critique Of Medicinal Conspicuousness Of Parsley (*Petroselinum Crispum*): A Culinary Herb Of Mediterranean Region. Pak J Pharm. Sci , 27(1):193-202.
- 30- Marinova, D; Ribarova , F. And Atanassova, M. (2005).Total Phenolics And Total Flavonoids In Bulgarian Fruits And Vegetables .J. Of The University Of Chem Technol Metallurgy , 40(3): 255-260.
- 31- Marzouk, M.; Soliman ,A. M. And Omar, T.Y. (2013). Hypoglycemic And Antioxidative Effects Of Fenugreek And Termis Seeds Powder In Streptozotocin-Diabetic Rats. European Rev Med Pharmacol Sci ., 17:559-565.
- 32- Meera, S; Gupta, V.S.S.S; Kumar, N.S.(2008). Immunomodulatory And Antioxidant Activity On A Polyherbal Formulation . Int. J Pharmacol, 4(4): 287-291.
- 33- Meyer, H; Bolannwa ,A; Wolfarm, G; And Linseisen, J. (2006). Bioavailability Of Apigenin From Apiin-Rich Parsley In Humans . Annals Of Nutr. Metab., 50: 167-172.
- 34- Mitchell,A.E; Hong, Y.J; Koh, E;Barrett, D.M; Bryant, D.E; Dension R.F. And Kaffka ,S.(2007).Ten-Year Comparision Of The Influence Of Organic And Conventional Crop Management Practices On The Content Of Flavonoids In Tomatoes. J Agric Food Chem., 55(15):6159-6159.
- 35- Mitra, E.; Ghosh, A.K.; Ghosh . D.; Mukherjee, D.; Chattopadhyay, A. And Dutta, S.(2012). Protective Effect Of Aqueous Curry Leaf (*Murraya Koenigii*) Extract Against Cadmium-Induced Oxidative Stress In Rat Heart. Food Chem Toxicol ,50:1340-53.
- 36- Navas-Acien, A; Aelvin, E; Sharrett, R; Calderon-Aeanda, E; Silbergeld, E. And Guallar, E. (2004). Lead ,Cadmium, Smoking, And Increase Risk Of Peripheral Arterial Disease . Circulation,109: 1-6.
- 37- Ognjanovic, B.I.; Markovic, S.D.; Pavloic, S.Z; Zikic, R.V; Stajn, .S. And Saicic, Z.S.(2008). Effect Of Chronic Cadmium Exposure On Antioxidant Defense System In Some Tissue Of Rats: Protective Effect Of Selenium. Physiol Res., 57:403-411.
- 38- Olisekodiaka MJ, Igbeneghu CA, Onuegbu AJ, Oduru R, Lawal AO. (2012). Lipid, Lipoproteins, Total Antioxidant Status And Organ Changes In Rats Administered High Doses Of Cadmium Chloride. Med Princi Pract.;21:156–159.
- 39- Oyewole, O.I.; Shoremi, M.O.; And Oladele, J.O.((2016). Modulatory Effects Of *Ricinus Communis* Leaf Extract On Cadmium Chloride-Induced Hyper-

- lipidemia And Pancytopenia In Rats. *Am J Biomedl Res.*, 4, (2):38-41. DOI:10.12691/Ajbr-4-2-2.
- 40- Oyinloye, B.E.; Ajiboye B.O.; Ojo, O. A.; Nwozo, S.O. And Kappo ,A.P. (2016). Cardioprotective And Antioxidant Influence Of Aqueous Extracts From *Sesamum Indicum* Seeds On Oxidative Stress Induced By Cadmium In Wistar Rats. *Pharmacol Mag.*,12 (Supl 2):S170-4. Doi: 10.4103/0973-1296.182155.
- 41- Ozturk, M; Buykakilli, B; Balli, E; Cimen, B; Gunes, S. And Erdogan, S. (2009). Determination Of Acute And Chronic Effects Of Cadmium On The Cardiovascular System Of Rats. *Toxicol Mechan Methods*, 19(4): 308-317.
- 42- Pápay, Z.E. And Antal, I.(2014). Study On The Antioxidant Activity During The Formulation Of Biological Active Ingredients. *European Sci J .*, 3 E - ISSN 1857- 7431 252.
- 43- Papuc, C.; Predescu, C.; Nicorescu, V.; Stefan, G.; And Nicorescu, I.(2016). Antioxidant Properties Of A Parsley (*Petroselinum Crispum*) Juice Rich In Polyphenols And Nitrites. 1st International Multidisciplinary Conference On Nutraceuticals And Functional Foods. *Curr Res Nutri Food Sci.*, 4(SI. 2), 114-118.
- 44- Prabu, S. M., Shagirtha, K. And Renugadevi, J. (2010), Amelioration Of Cadmium-Induced Oxidative Stress, Impairment In Lipids And Plasma Lipoproteins By The Combined Treatment With Quercetin And α -Tocopherol In Rats. *J Food Sci.*, 75: T132–T140.
- 45- Prabu, S.M.; Muthumani, M. And Shagirtha, K.(2013). Quercetin Potentially Attenuates Cadmium Induced Oxidative Stress Mediated Cardiotoxicity And Dyslipidemia In Rats. *European Rev Med Pharmacol Sci*,17:582-95.
- 46- Peters, J; Perlstein, T; Perry, M; Mcneely, E. And Weuve, J. (2010). Cadmium Exposure In Association With History Of Stroke And Heart Failure; *Environmental Research*,110:199-206.
- 47- Rogalska, J., Brzóška, M.M., Roszczenko, A., Moniuszko- Jakoniuk, J., 2009. Enhanced Zinc Consumption Prevents Cadmium-Induced Alterations In Lipid Metabolism In Male Rats. *Chem. Biol. Interact.*, 177(2):142-152.
- 48- Salman, I.M. And Inamdar, N.(2012). Effect Of Gliclazide On Cardiovascular Risk Factors Involved In Split Dose Streptozotocin Induced Neonatal Rat Model: A Chronic Study. *International Journal Of Basic & Clinical Pharmacology.*, 1(3):196-201.
- 49- Samarghandian, S.; Azimi-Nezhad, M.; Shabestari, M.M.; Azad, F.J.; Farikhondeh, T. And Bafandeh, F. (2015). Effect Of Chronic Exposure To Cadmium On Serum Lipid, Lipoprotein And Oxidative Stress Indices In Male Rats. *Interdiscip Toxicol.* 2015 Sep; 8(3): 151–154. Doi: 10.1515/Intox-2015-0023.
- 50- Satarug, S; Haswell, E. And Moore, M.R. (2000). Safe Levels Of Cadmium Intake To Prevent Renal Toxicity. *Br J Nutr.* 84(6):802-971.

- 51- Satarug, S; Nishijo, M; Jerome, M; Lasker, M. Robert, J; Edwards, M. And Moore, R.(2006).Kidney Dysfunction And Hypertension :Role Fo Cadmium ,P450 And Heme Oxygenases. The Tohoku J. Exp. Med, 208(3): 179-202.
- 52- Scalbert, A; Manach, C; Manach, C; Morand. C; Remesy, C; And Jimenez, L. (2005). Dietary Polyphenols And The Prevention Of Disease . Crit.Rev.Food Sci .Nutr., 45(4): 287-306.
- 53- Sharma,Y.; Nagar,A.; Srivastava, N.S. And Bala, K.(2017). Antioxidant Activity Of Polyphenolic Flavonoid Of Stem Of Nicotiana Tabacum. American Journal Of Drug Discovery And Development Year: 2017 | Volume: 7 | Issue: 1 | Page No.: 25-32.
- 54- Shuka, R. And Kumar, M.,(2009). Role Of Panax Ginseng As An Antioxidant After Cadmium Induced Hepatic Injury .Food Chem. Toxicol., 47(4): 769-773.
- 55- Snedecor, G.W. And Cochran ,W.G. (1973).Statistical Methods. 6th The Iowa State University Press.,: 2380248.
- 56- Soliman,H.A.; Eltablawy,N.A. And Hamed, M.S. .(2015). The Ameliorative Effect Of *Petroselinum Crispum* (Parsley) On Some Diabetes Complications. J Medicinal Plants Studies., 3(4): 92-100.
- 57- Soliman, H.A.; El-Desouky, M.A.; Hozayen, W.G.; Ahmed, R.R. And Khaliefa, A.K. (2016). Hepatoprotective Effects Of Parsley, Basil, And Chicory Aqueous Extracts Against Dexamethasone-Induced In Experimental Rats. J Intercult Ethnopharmacol. 5(1): 65–71.
- 58- Ververidis ,F; Trantas, E; Douglas, C; Vollmer, G; Kretzschmar, G. And Panopoulos, N. (2007). Biotechnology Of Flavonoids And Other Phenylpropanoid- Derived Natural Products. Part I: Chemical Diversity, Impacts On Plant Biology And Human Health .Biotechnol J.2 (10): 1214-34.
- 59- Young, C.E.; Karas, R.H. And Kuvin, J.T.(2004). High Density Lipoprotein Cholesterol And Coronary Heart Disease. Cardio Rev., 12:107-119.
- 60- Yousuf ,H.A.; Al-Zubaidi, F.S. And Yousif, W.H.(2014). Study Of The Interaction Effect Between Parsley *Petroselinum Crispum* And Cadmium On Lipid Profile, Lipid Peroxidation And Catalase Activity Of Albino Mice Males' Liver And Kidney. Iraqi J Sci. 55:711–21.