

Available at https://www.iasj.net/iasj

Iraqi Academic Scientific Journals

Journal homepage: https://journals.uokerbala.edu.iq/index.php/UOKJ



### **Research Article**

# The effect of Different concentrations of glucose solution on the growth and biofilm formation of sensitive and resistant Pseudomonas aeroginosa isolates.

Suhad Hadi Mohammed, Hadi Mahmoud Hadi, Noor Al-Huda Kareem Hamed, Noor Kareem Abbas Department of Clinical laboratories, College of Applied Medical Sciences, University of Kerbala

Article Info	Abstract
Article history:	Burns are a severely debilitating class of wound. <i>Pseudomonas aureugnosa</i> is one of most common bacteria that associated with burn infection. Hypertonic glucose solution considered as a new approach to control chronic wound infections. To study the effect of glucose solution (at different concentrations) on the growth of <i>P. aeruginosa</i> and on their
Received 31-10-2022	
Received in revised	biofilm formation (for both resistant and sensitive isolates), Four <i>P</i> .
form 22-11-2022	aeruginosa isolates were isolated from swab samples taken from different sites of burn's patients whom admitted to Al-Imam Al-Hussain Medical
Accepted 6-12-2022	City. Antibiotic susceptibility patterns were determined and the effect of different concentration of glucose solution on growth and biofilm
Available online 06-06-	formation were studied.
2022	The results showed that the growth of all isolates (resistant and sensitive) were reduced for more than 50% after 5 hours of incubation and this
Keywords: Burns,	reduction increases both with the increasing of glucose concentration and
Pseudomonas	with the increasing of time of exposure of the bacterial isolates to the glucose solution. The biofilm formation was increased in the presence of
aerogenosa, Hypertonic	glucose solution with the used concentrations with the exception of the concentration of 100 mg in which the biofilm formation was inhibited in
solution, Biofilm,	comparison to controls. Regarding Biofilm eradication assay, the formed
Growth rate	biofilm layer was partially eradicated in the glucose concentrations of 50, 100 in most of the isolates.
	In conclusion, Hypertonic glucose solution in different concentrations has inhibitory effect (both concentrations dependent and time dependent) on growth of sensitive and resistant bacteria. The inhibitory action of the glucose against <i>P. aeruginosa</i> biofilms was found in the lower concentration of glucose solution whereas higher concentration of glucose increased the biofilm formation. none of the concentrations revealed the

Corresponding Author E-mail :

Peer review under responsibility of Iraqi Academic Scientific Journal and University of Kerbala.

complete destruction of biofilm.

#### Introduction

Burns are considered a notorious type of wound which could have a negative effect on patient's life. The annual number of deaths due to fire, heat and hot substance is about 300,000 (1). Higher frequency of Burns that result from fire are recorded in less developing countries (2–5). Damaging of tissue at burn site leads to the loss of the biological fluids which defined as burn wound exudates (6). The microenvironment of these burn wound excaudate will provide a good environment to certain pathogens to proliferate successfully.

Infection of the wound might results in approximately 75% of cases to death. Gramnegative bacteria like *Pseudomonas aeruginosa*, *Acinetobacter baumannii*, *Klebsiella* spp., *Stenotrophomon as* spp., *Escherichia coli*, and *Enterobacter cloacae* are frequently isolated from wounds. These pathogens are responsible for most of the nosocomial infections and responsible for outbreaks in burn wards globally (7–9).

*Pseudomonas aereugnosa* is an opportunistic pathogen associated with burn infections and is responsible for a variety of acute and chronic infections including endocarditis, pneumonia, and infections of the urinary tract, central nervous system, wounds, eyes, ears, skin, and musculoskeletal system (10,11) due to its highly resistance rates to antibiotics

#### Material & Methods Bacterial isolates

Four *P. aeruginosa* isolates were isolated from swab samples taken from different sites of burn's patients whom admitted to Al-Imam Al-Hussain Medical City during the period of sample collection (two months, from Nov.

#### Antibiotic Susceptibility patterns

Susceptibility patterns of the isolated bacteria was done using VITEK2 system for gram negative bacteria. Four isolates were selected based on susceptibility test recovered from VITEK2 system, two isolates were sensitive and two other isolates were resistant to most of the tested antibiotics. MIC of the antimicrobial agents were tested within (12). The fast increase in drug resistance among bacteria has become a serious health problem health worldwide. To overcome this increase, broad-spectrum antibiotics have been used which leads to the selection and accumulation of resistance strains of bacteria (13,14).

*Pseudomonas aereugenosa* is found in different environmental due to its simple requirements for growth. Formation of biofilms in addition to the survival of this bacteria on both living and non-living surfaces are considered the main causes for burn wound complication (15,16). Biofilm is defined as aggregations of one or more than one type of bacterial species that found in an extracellular matrix which helps the bacteria to survive in their hostile environment.

There are many solutions for treatment of burns but the most effective of them is glucose solution. Dressings of the wound with hypertonic glucose solution could be used as an alternative method in wound management which enhance the auto-debridement process, increase proliferation of granulation, and reduce the patient's healing time (17,18).

Thus, the current study Aims to study the inhibitory effect of glucose solution (at different concentrations) on growth and biofilm formation of resistant and sensitive isolates of *P. aeruginosa*.

2021 to Jan 2022) and these isolates were used in the following experiments after identification of the bacteria using VITEK 2 compact system. All isolates were subcultured in Blood and MacConkey agar before it used to ensure its activity and purity.

VITEK2 system including Amoxicillin, Amoxicillin-Clavulanic acid, Ampicillin/Sulbactam, Ticarcillin, Ticarcillin/Clavulanic Acid, levofloxacin, Amikacin. Gentamicin. Meropenem, Norfloxacin, Imipenem, Ciprofloxacin, Ofloxacin, Tobramycin, colistin, Piperacillin, Ciprofloxacin, Cefepime, Tetracycline.

#### **Glucose solution**

Seven concentrations of glucose solution (D-(+)-glucose, Anhui Herrman Impex Co., Ltd)

# The effect of hypertonic glucose solution on growth of *P. aeruginosa*

The effect of hypertonic glucose solution was done using the method prescribed previously (19). Briefly, 50ul of overnight bacterial suspension  $(10^8)$  were added to 4950ul (Final volume is 5 ml ) nutrient broth media to prepare 1% (v/v) dilution for each isolate.

#### **Biofilm Inhibition test**

Biofilm formation inhibition test was done using the method prescribed previously (19). Briefly, 100ul of overnight bacterial solution with nutrient broth (50ul of bacteria suspension ( $10^8$  CFU) was mixed with 4950ul nutrient broth media to prepare 1% v/v dilution) and 100ul of different concentration of glucose were added into two wells for each concentration and for each isolate. Then, the plate were incubated for 24h to allow the

#### **Biofilm Eradication test**

Biofilm formation inhibition test was assessed using method described previously (19). Briefly, 100ul of overnight bacterial solution with nutrient broth (1% v/v) was added to each well and incubated at 37 °c for 24 hours. The following day, 100ul of glucose solution was added to each well (each concentration repeated twice) and 100ul distilled water was added to control wells. After incubation for

#### **Ethical statement:**

The current study was approved by the scientific committee in Department of Clinical Laboratories in College of Applied Medical Sciences, Kerbala University. The

#### **Statistical Analysis:**

The mean of the absorbance reading was calculated and No other statistical analysis

#### **Results:**

In the current study, four *Pseudomonas aeruginosa*, were isolated from wound samples taken from different sites of burn's

were prepared (50 ,100 ,200 ,300 ,400 ,500, 600 mg/ml) to be used.

One hundred microliter of the bacterial solution were added to each well in the microtiter plate. Then, 100ul of glucose solution (each concentration was repeated twice) was added to each well. A hundred microliter of nutrient broth was added as a negative control. After 5 h, 8 h, and 24 h of incubation at 37°C, the absorbance were measured by ELISA reader (PKL PPC 142) at wave length 630 nm.

bacteria to grow and form Biofilm. The following day, the mixture was removed from the wells and the attached bacteria was washed. One percent Crystal Violate (CV) solution (200ul) was added to each well for 20 min followed by washing step. The absorbed CV dye were eluted using absolute Ethanol after addition of 200ul to each well and the absorbance were read immediately by using ELISA reader (OD = 630nm).

24h, The mixture was removed from the wells and the attached cells was rinsed. Crystal Violate (200ul of 1% CV) solution was added to each well for 20 min followed by washing step. The absorbed dye was eluted using absolute Ethanol (200ul were added to each well) and the absorbance were read immediately by using ELISA reader (OD = 630nm).

study design didn't include any patients and is based on laboratory work only on bacterial isolates, thus , No ethical consent was required.

was used. Figures were constructed using Excel program.

patients. The identification and antibiotic susceptibility testing were performed using VITEK2 compact system. The selection of these isolates were done based on the results of susceptibility testing. Thus, two isolates which are sensitive and two other isolates

# The effect of hypertonic solution on bacterial growth.

The results showed that the growth of all isolates (resistant and sensitive) were reduced for more than 50% after 5 hours of incubation

which are resistant to most of the tested antibiotics were enrolled.

and this reduction increases both with the increasing of glucose concentration and with the increasing of time of exposure of the bacterial isolates to the glucose solution(after 8hrs and24hrs), as demonstrated in figures 1-4.

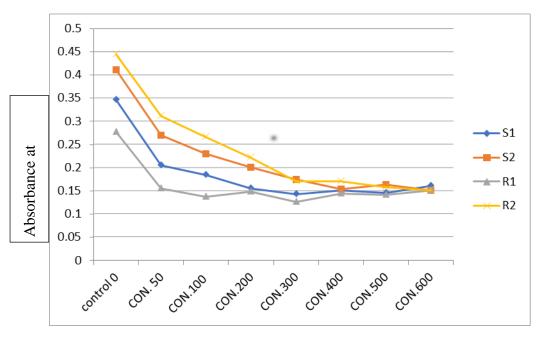


Figure 1. Growth of bacterial isolates in different concentrations of glucose solution after 5 hours of incubation

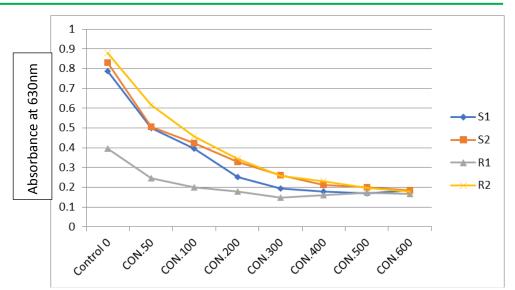


Figure 2. Growth of bacterial isolates in different concentrations of glucose solution after 8 hours of incubation

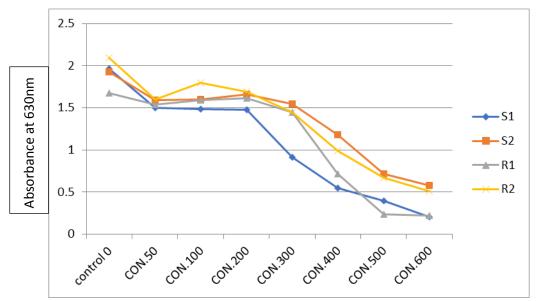


Figure 3. Growth of bacterial isolates in different concentrations of glucose solution after 24 hours of incubation

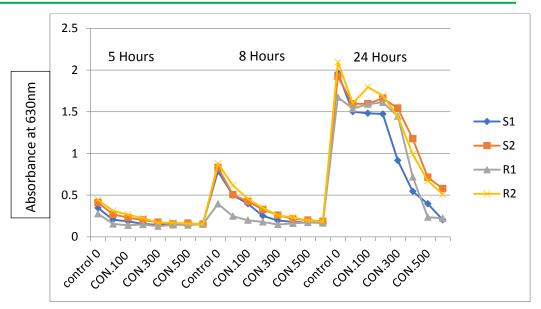


Figure 4: Growth of *Pseudomonas aerogenosa*. With the different glucose concentration for 5, 8, 24 hours of incubation.

**Effect of hypertonic solution on formation and eradication of Biofilm** The antibiofilm activity of glucose solution were measured by using crystal violet assay. The current study found that the biofilm formation was increased in the presence of glucose solution with the used concentrations with the exception of the concentration of 100 mg in which the biofilm formation was inhibited in comparison to controls, as shown in figure 5. Regarding Biofilm eradication assay, the formed biofilm layer were eradicated in the glucose concentrations of 50, 100, and 600 mg as shown in Figure 6.

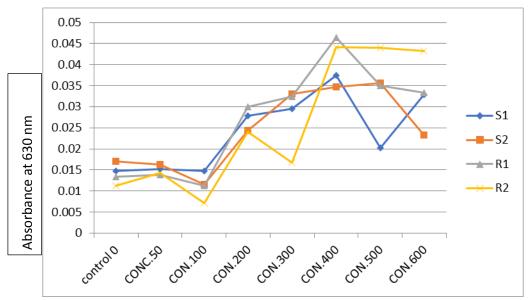


Figure 5. Biofilm formation inhibition Assay

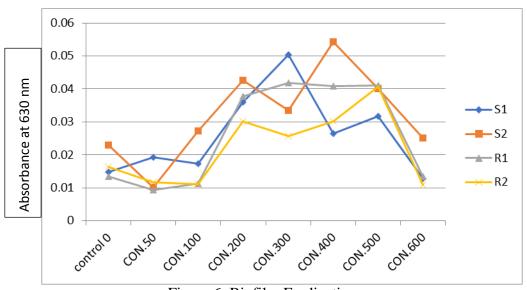


Figure 6. Biofilm Eradication assay

# Discussion

It has been documented that burns considered as sever forms of trauma that need special medical care. The microenvironment of burn wounds are good environment for bacterial growth and proliferation (20,21).

*Pseudomonas aeruginosa*, is one of the most commonly isolated pathogen from infected burns with high resistance rate to most of antibiotics and subsequently alternative methods for burn management is required (22). Additionally, this bacteria characterized by formation of biofilm which considered as an obstacle for treatment of wound infection because bacteria within biofilm are protected from immune response and antibiotics (23).

Although treatment of burn wounds with glucose solution has been used previously (24), the inhibitory effect of this solution on bacterial growth and biofilm formation ability has not documented. This study investigated the inhibitory action on bacterial growth and biofilm formation on resistant and sensitive bacterial isolates.

The current study revealed that hypertonic glucose solution in different concentrations has inhibitory effect (both concentrations dependent and time dependent) on growth of sensitive and resistant bacteria. Similar findings were documented by previous study (25).

It has been documented that Pseudomonas has the ability to form biofilm on different medical surfaces. Bacteria inside biofilm are much more resistant to antimicrobial agents than planktonic forms since bacteria that are unresisting to antimicrobial agents in any way can turn resistant after forming a biofilm(26). This biological development protects the pathogen from host immunity and contributes to its antimicrobial resistance. It is estimated that about 80% of infectious diseases are due to biofilm formation. Biofilm-forming ability and antimicrobial resistance of this pathogen lead to many persistent and chronic bacterial infections) (27,28).

It is the most commonly isolated species from chronic wounds and is considered a potent biofilm producer since they act as a barrier in wound healing and exhibits high resistance to antimicrobial therapy (29,30). Thus, We further explored the effect of glucose solution on biofilm for this bacteria. To illustrate this aim, the antibiofilm activity of various concentration of glucose were tested crystal violet staining technique. The results revealed that The inhibitory effects of the glucose towards P. aeruginosa biofilms was found in the lower concentration of glucose solution for 75% of the isolates. However, higher concentration of glucose result in higher level of biofilm formation. Similar finding was documented by Wang et al., who reported that glucose induce biofilm formation (24).

Concerning Eradication assay, the used concentrations didn't reflect complete eradication of biofilm. However, glucose concentration of 100 mg/mL considered the best concentration which might possibly reflect the inhibitory action of glucose solution on expression of the genes involved in the biofilm formation. Similar findings was

## **Conclusion:**

Hypertonic glucose solution in different concentrations has inhibitory effect (both concentrations dependent and time dependent) on growth of sensitive and

# References

- WHO. No Title Global Burden of Disease Collaborative Network. Global Burden of Disease Study 2019 (GBD 2019) Reference Life Table. Seattle, United States of America: Institute for Health Metrics and Evaluation (IHME), 2021. DOI https://doi.org/10.6069/1D4Y-YQ3. Glob Heal Data Exch. 2021;
- Chichom Mefire A, Fokou M. Epidemiology of paediatric injury in low income environment: value of hospital based data prior to the institution of a formal registration system. Afr J Paediatr Surg. 2013;10(3):265–70.
- Chichom-Mefire A, Mbarga-Essim NT, Monono ME, Ngowe MN. Compliance of district hospitals in the Center Region of Cameroon with WHO/IATSIC guidelines for the care of the injured: a cross-sectional analysis. World J Surg. 2014 Oct;38(10):2525–33.
- Guest JF, Fuller GW, Edwards J. Cohort study evaluating management of burns in the community in clinical practice in the UK: Costs and outcomes. BMJ Open. 2020;10(4):1–12.
- Smolle C, Cambiaso-Daniel J, Forbes AA, Wurzer P, Hundeshagen G, Branski LK, et al. Recent trends in burn epidemiology worldwide: A systematic review. Burns. 2017 Mar;43(2):249–57.
- 6. Cutting KF. Wound exudate: composition and functions. Br J Community Nurs. 2003;8(Sup3):S4–9.
- 7. Williams FN, Lee JO. Pediatric Burn Infection. Surg Infect (Larchmt). 2021;22(1):54–7.
- 8. Ramakrishnan M, Putli Bai S, Babu M. Study on biofilm formation in burn wound infection in a pediatric hospital in Chennai, India. Ann Burns Fire Disasters [Internet].

reported by Chen et al., (25).

resistant bacteria. Lower concentration of glucose solution is required for biofilm formation inhibition and higher concentration of glucose increased the biofilm formation. Eradication of biofilm formation does not occur with all concentration.

2016;29(4):276–80. Available from: http://www.ncbi.nlm.nih.gov/pubmed/2828 9362%0Ahttp://www.pubmedcentral.nih.g ov/articlerender.fcgi?artid=PMC5347310

- Branski LK, Al-Mousawi A, Rivero H, Jeschke MG, Sanford AP, Herndon DN. Emerging infections in burns. Surg Infect (Larchmt). 2009;10(5):389–97.
- Genetics a n d Biochemistry of Pseudomonas By Pierre Louisot . 1975;3(3):1975.
- Rocha CL, Coburn J, Rucks EA, 11. Olson JC. Characterization of Pseudomonas aeruginosa exoenzyme S as bifunctional enzyme in J774a.1 а macrophages. Infect Immun. 2003;71(9):5296-305.
- 12. Ak O, Batirel A, Ozer S, Çolakoğlu S. Nosocomial infections and risk factors in the intensive care unit of a teaching and research hospital: a prospective cohort study. Med Sci Monit Int Med J Exp Clin Res. 2011 May;17(5):PH29-34.
- Bassetti M, De Waele JJ, Eggimann P, Garnacho-Montero J, Kahlmeter G, Menichetti F, et al. Preventive and therapeutic strategies in critically ill patients with highly resistant bacteria. Intensive Care Med. 2015 May;41(5):776– 95.
- 14. Al-Orphaly M, Hadi HA, Eltayeb FK, Al-Hail H, Samuel BG, Sultan AA, et al. Epidemiology of Multidrug-Resistant Pseudomonas aeruginosa in the Middle East and North Africa Region. mSphere. 2021 May;6(3).
- 15. Flemming H-C, Wingender J, Szewzyk U, Steinberg P, Rice SA, Kjelleberg S. Biofilms: an emergent form of bacterial life. Nat Rev Microbiol [Internet]. 2016;14(9):563–75. Available from:

https://doi.org/10.1038/nrmicro.2016.94

- 16. Longo F, Vuotto C, Donelli G. Longo 2014. New Microbiol. 2014;37:119–27.
- Mangete ED, West KS, Blankson CD. Hypertonic saline solution: an effective wound dressing solution. East Afr Med J. 1993 Feb;70(2):104–6.
- Rabago D, Zgierska A, Fortney L, Kijowski R, Mundt M, Ryan M, et al. Hypertonic Dextrose Injections (Prolotherapy) for Knee Osteoarthritis: Results of a Single-Arm Uncontrolled Study with 1-Year Follow-Up. J Altern Complement Med [Internet]. 2012 Apr 1;18(4):408–14. Available from: https://doi.org/10.1089/acm.2011.0030
- 19. She P, Wang Y, Luo Z, Chen L, Tan R, Wang Y, et al. Meloxicam inhibits biofilm formation and enhances antimicrobial agents efficacy by Pseudomonas aeruginosa. Microbiologyopen. 2018;7(1):1–9.
- 20. Brown SA, Palmer KL, Whiteley M. Revisiting the host as a growth medium. Nat Rev Microbiol. 2008;6(9):657–66.
- 21. Jones EM, Cochrane CA, Percival SL. The Effect of pH on the Extracellular Matrix and Biofilms. Adv Wound Care. 2015;4(7):431–9.
- 22. Fournier A, Voirol P, Krähenbühl M, Bonnemain C-L, Fournier C, Pantet O, et al. Antibiotic consumption to detect epidemics of Pseudomonas aeruginosa in a burn centre: A paradigm shift in the epidemiological surveillance of Pseudomonas aeruginosa nosocomial infections. Burns. 2016 May;42(3):564–70.
- Maslova E, Eisaiankhongi L, Sjöberg F, McCarthy RR. Burns and biofilms: priority pathogens and in vivo models. npj Biofilms Microbiomes [Internet]. 2021;7(1):73. Available from: https://doi.org/10.1038/s41522-021-00243-2
- 24. She P, Wang Y, Liu Y, Tan F, Chen L, Luo Z, et al. Effects of exogenous glucose on Pseudomonas aeruginosa biofilm formation and antibiotic resistance. Microbiologyopen. 2019;8(12):1–15.
- 25. Chen T, Xu Y, Xu W, Liao W, Xu C, Zhang X, et al. Hypertonic glucose inhibits growth and attenuates virulence factors of multidrug-resistant Pseudomonas

aeruginosa. BMC Microbiol [Internet]. 2020;20(1). Available from: https://doi.org/10.1186/s12866-020-01889-2

- 26. Dincer S, Uslu FM, Delik A. Antibiotic Resistance in Biofilm. In: Dincer S, Özdenefe MS, Arkut A, editors. Bacterial Biofilms [Internet]. Rijeka: IntechOpen; 2020. Available from: https://doi.org/10.5772/intechopen.92388
- 27. Elmanama AA, Al-Sheboul S, Abu-Dan RI. Antimicrobial Resistance and Biofilm Formation of Pseudomonas aeruginosa. Int Arab J Antimicrob Agents. 2020;10(2):1–11.
- 28. Lewis K. Riddle of biofilm resistance. Antimicrob Agents Chemother. 2001;45(4):999–1007.
- 29. Mantero M, Gramegna A, Pizzamiglio G, D'Adda A, Tarsia P, Blasi F. Once daily aerosolised tobramycin in adult patients with cystic fibrosis in the management of Pseudomonas aeruginosa chronic infection. Multidiscip Respir Med [Internet]. 2017;12(1):1–4. Available from: http://dx.doi.org/10.1186/s40248-016-0083-y
- 30. Davis R, Brown PD. Multiple antibiotic resistance index, fitness and virulence potential in respiratory Pseudomonas aeruginosa from Jamaica. J Med Microbiol. 2016;65(4):261–71.